

ANTIOXIDANT POTENTIAL AND SOME MINERAL CONTENTS OF WILD EDIBLE MUSHROOM *Ramaria stricta*

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Abstract

People have used mushrooms for many different diseases for thousands of years. Although mushrooms are considered as nutrients for the first time, it has been determined by researches that they have medicinal properties. In this study, antioxidant status, oxidant status and some mineral contents of wild edible mushroom *Ramaria stricta* (Pers.) Quél were determined. The antioxidant and oxidant states of the mushroom were measured using Rel Assay kits. Element contents were determined using atomic absorption spectrometry. In this study, the antioxidant potential of *R. stricta* mushroom used in our study has been evaluated to be high in term of total antioxidant status (TAS) values (4.223 ± 0.054 mmol/l), total oxidant status (TOS) values (8.201 ± 0.095 μ mol/l), and oxidative stress index (OSI) value (0.194 ± 0.001). Fruitbodies of *R. stricta* contains essential mineral elements which are of immense health benefit. The highest Fe content (451.21 ± 5.56 mg.kg⁻¹) was found in mushroom samples. It was established high Cu content (95.54 ± 2.06 mg.kg⁻¹), and Zn content (39.19 ± 1.07 mg.kg⁻¹). Ni content was 7.17 ± 0.32 mg.kg⁻¹. A lower content (2.18 ± 0.10 mg.kg⁻¹) was recorded for Pb. In addition, element contents were found to be at normal levels. As a result, *R. stricta* mushroom is thought to be a natural antioxidant source.

Key words: antioxidant, oxidant, edible mushroom, *Ramaria stricta*.

INTRODUCTION

Mushrooms are one of the most important elements of natural ecosystems. They are the indispensable elements of the ecological cycle. They are involved in the disintegration of organic matter in environments where they spread (Canli et al., 2016; İnci and Kırbag, 2018). Besides their ecological importance, mushrooms are important natural foods. It is one of the delicious foods of the human diet. They are rich in minerals (Gürgen et al., 2018). They also contain many essential amino acids, proteins, being rich in vitamin B. Edible mushrooms proved significant medicinal properties due to their high polysaccharide content, especially β -glucans (İnci et al., 2019). Many studies have shown that mushrooms have different medicinal properties (Gedik et al., 2019). In previous studies, mushrooms have been reported to have many different activities such as antioxidant, antimicrobial, anti-cancer, anti-angiogenic, anti-inflammatory, anti-proliferative, antitumor, anti-HIV, anti-

genotoxic, anti-aging, anti-gout, anti-cholinesterase, anti-hyperlipemia, anti-gastric ulcer activity, anti-allergy, DNA protective activity, anti-hypoxic, anti-diabetic, Immunomodulatory activity and anti-thrombotic activity (de Oliveira et al., 2002; Jose et al., 2002; Song et al., 2003; Kim et al., 2004; Lavi et al., 2006; Bae et al., 2007; El Dine et al., 2008; Weng et al., 2010; Kang et al., 2011; Kim et al., 2013; Li et al., 2015; Pandimeena et al., 2015; Devi and Maiti, 2016; Nguyen et al., 2016; Bal et al., 2017; Sevindik et al., 2018a).

Ramaria is a genus with about 200 species. There are species that spread in different ecosystems of the world of *Ramaria* genus. In addition to *R. formosa* and *R. pallida*, which cause nausea, vomiting and diarrhea, there are also types such as edible *R. flava*. The antioxidant activities of some *Ramaria* species are shown in Table 1. *Ramaria stricta* (Pers.) Quél., commonly known as the strict-branch coral of the genus *Ramaria* (Petersen and Scates, 1988; Braeuer et al., 2018). It has a

cosmopolitan distribution, and grows on dead wood, stumps, trunks, and branches of both deciduous and coniferous trees (Kuo, 2009).

Table 1. Antioxidant activities of *Ramaria* species

Mushroom species	Country	Extraction	References
<i>Ramaria flava</i>	China, Turkey	Ethanol, Hexane, Methanol, Aqueous	Gursoy et al., 2010; Liu et al., 2013; Öztürk et al., 2014; Bozdogan et al., 2016; Sadi et al., 2016
<i>R. largentii</i>	Romania	Ethanol	Aprotosoae et al., 2017
<i>R. botrytis</i>	China	Water, Methanol, petroleum ether	Kim and Li, 2003; Froufe et al., 2009; Li 2017
<i>R. aurea</i>	India	Ethanol, methanol, Aqueous	Khatua et al., 2015; Zengin et al., 2017
<i>R. formosa</i>	India	Methanol	Ramesh and Pattar, 2010
<i>R. botrytoides</i>	China	Methanol	Guo et al., 2012
<i>R. subalpina</i>	India	Methanol	Acharya et al., 2017
<i>R. fennica</i>	Turkey	Methanol	Bakır et al., 2018

The aim of this study is the determination of antioxidant status, oxidant status and Fe, Zn, Cu, Pb and Ni contents of wild edible mushroom *Ramaria stricta* (Pers.) Quéf.

MATERIALS AND METHODS

R. stricta samples were collected in 2019 from Bilyayivka, Odessa region (Ukraine). The mushroom samples were dried in an incubator at 40°C. 30 g of dried mushroom samples were weighed and powdered. Then, mushroom was extracted with ethanol (EtOH) in the Soxhlet apparatus during approximately 6 hours at 50°C. A rotary evaporator was used to remove the solvent from the obtained extract. The extracts were kept at +4°C until experiment.

Total Antioxidant and Oxidant Studies

Total antioxidant status (TAS) values were determined using Rel Assay TAS kits (Erel, 2004). Total oxidant status (TOS) values were determined using Rel Assay TOS kits (Erel, 2005). Trolox was used as TAS calibrator and hydrogen peroxide was used as TOS calibrator. Oxidative stress index (OSI) value (Arbitrary unit: AU) was determined according to the formula below (Erel, 2005):

$$\text{OSI (AU)} = \frac{\text{TOS, } \mu\text{mol H}_2\text{O}_2 \text{ equiv./l}}{\text{TAS, mmol Trolox equiv./l} \times 10}$$

Determination of Element Content

Samples were dried at 80°C until constant weight to determine the Fe, Zn, Cu, Pb, and Ni contents of *R. stricta*. It was mineralized in a mixture of 9 ml HNO₃+1 ml H₂O₂ in the microwave solubilizer (Milestone Ethos Easy) by taking 0.5 g of 5 samples. The element contents of the samples were determined using an atomic absorption spectrophotometer device (Agilent 240FS AA) (Sevindik et al., 2017).

Statistical analysis

The experimental results were expressed as means ± SEM (standard error of the mean) of triplicates. The data was analysed with Excel statistical functions using the Microsoft Office 2016. Differences at P ≤ 0.05 were considered to be significant.

RESULTS AND DISCUSSIONS

Total Antioxidant and Oxidant Status

Mushrooms have the potential to produce many antioxidant enzymes and coenzymes in secondary metabolite structure. Mushrooms, which are rich in nutrient content, contain vitamins with strong antioxidant characteristics such as A, C and E. In this context, it is very important natural products (Rathore et al., 2017). TAS values are an indicator of the endogenous antioxidants they produce (Selamoglu et al., 2016). Mushrooms with high TAS values have the potential to be an important antioxidant natural source. TOS values are an indicator of produced oxidant compounds (Selamoglu et al., 2017). High TOS values show that mushrooms produce more reactive oxygen types under the influence of environmental factors and people should pay attention to the consumption of these products. Thus, these indicators allow us to evaluate the antioxidant potential in the complex. However, there are very few studies to determine TAS, TOS and OSI values of mushrooms. In our study, TAS, TOS and OSI values of EtOH extract of *R. stricta* were determined. The findings obtained are shown in Table 2.

In literature, TAS, TOS and OSI values of *R. stricta* have not been determined. But, there are analogical studies of the antioxidant potential of different wild mushrooms.

Table 2. TAS, TOS and OSI values of *R. stricta*

Sample	TAS (mmol/l)	TOS ($\mu\text{mol/l}$)	OSI
<i>R. stricta</i>	4.223 \pm 0.054	8.201 \pm 0.095	0.194 \pm 0.001

*Values are presented as mean \pm S.D

*Experiments were made as 5 parallel

Compared to these studies, TAS value of *R. stricta* was found higher than *Cerioporus varius* (2.312 mmol/l) (Sevindik, 2019), *Laetiporus sulphureus* (2.195 mmol/L) (Sevindik et al., 2018b), *Lepista nuda* (3.102 mmol/l) (Bal et al., 2019), *Lentinus tigrinus* (1.748 mmol/l) (Sevindik, 2018), and lower than *Leucoagaricus leucothites* (8.291 mmol/l) (Sevindik et al., 2018c). It is thought that this difference between the TAS values of mushrooms is due to the differences in their potential to produce antioxidant compounds. In addition, these differences may occur depending on the differences, levels, and diversity of secondary metabolites produced by the mushrooms as a reaction of the defense system depending on internal and external factors. The high TAS value of *R. stricta* in our study shows that mushrooms can be used as an important natural antioxidant source. In the literature, it was also reported that petroleum ether extracts of *R. stricta* show significant antioxidant activity (Sharma and Gautam, 2017).

Compared to TOS values of other mushroom species, it was established that TOS value of *R. stricta* was higher than those of *L. sulphureus* (1.303 $\mu\text{mol/l}$) (Sevindik et al., 2018b) and lower than *L. tigrinus* (19.294 $\mu\text{mol/l}$) (Sevindik, 2018), *C. varius* (14.358 $\mu\text{mol/l}$) (Sevindik, 2019), *L. nuda* (36.920 $\mu\text{mol/l}$) (Bal et al., 2019), and *L. leucothites* (10.797 $\mu\text{mol/l}$) (Sevindik et al., 2018c).

The differences in the TOS studies conducted with different mushroom species obtained from different localities attract attention.

It is thought that the main reason for this difference in TOS values is due to differences

in the environmental factors and metabolic processes of the mushrooms. Consumption of a natural product with a high TOS value may pose problems for human health (Korkmaz et al., 2018).

And according to our results, the OSI values of *R. stricta* samples had higher values than *L. leucothites* (0.130) (Sevindik et al., 2018c) and *L. sulphureus* (0.059) (Sevindik et al., 2018b), and lower than *L. tigrinus* (1.106) (Sevindik, 2018), *C. varius* (0.627) (Sevindik, 2019), and *L. nuda* (1.190) (Bal et al., 2019). These differences in OSI values arise from the fact that the antioxidant system of mushroom is effective at different levels against oxidant compounds. As a result, OSI values of *R. stricta* were low.

This is because the fungi antioxidant system is potent and effective against oxidant compounds.

Element Contents

Mushrooms, which acts as a decomposing role in the ecosystem, accumulate different levels of elements within the body depending on the substrate content. Because of these properties, they are important element indicators (Baba et al., 2012). In this study, Fe, Cu, Zn, Pb and Ni contents of *R. stricta* were determined. The results are shown in Table 3.

Element contents may vary depending on the habitats collected from mushrooms consumed in different parts of the world. The maximum and minimum ranges of Fe, Zn, Cu, Pb and Ni levels detected in different wild mushrooms were shown in Table 4.

It was observed that the contents of Fe, Zn, Cu and Pb of *R. stricta* used in our study were in line with literature data. Ni content of *R. stricta* was found to be higher than the ranges specified in the literature.

Table 3. Element contents of *R. stricta*

Sample	Fe (mg.kg ⁻¹)	Zn (mg.kg ⁻¹)	Cu (mg.kg ⁻¹)	Pb (mg.kg ⁻¹)	Ni (mg.kg ⁻¹)
<i>R. stricta</i>	451.21 \pm 5.56	39.19 \pm 1.07	95.54 \pm 2.06	2.18 \pm 0.10	7.17 \pm 0.32

Values are presented as mean \pm S.D.

n = 3 (Experiments were made as 3 parallel).

Table 4. Range of reported literature values (mg/kg dry weight) of different mushrooms

Elements	Values (mg/kg dry weight)	References
Iron (Fe)	14.60-1714	Sarikurkcü et al., 2011; Sevindik et al., 2018a
Zinc (Zn)	7.63-240	Falandysz et al., 2007
Copper (Cu)	1.90-180	Falandysz et al., 2008; Sevindik et al., 2018c
Lead (Pb)	0.18-16.54	Nikkarinena and Mertanen, 2004; Sevindik et al., 2018a
Nickel (Ni)	0.67-6.72	Vetter, 1990; Sevindik et al., 2018a

CONCLUSIONS

In this study, total antioxidant level, total oxidant level, oxidative stress index and element contents of wild edible mushroom *R. stricta* were determined. Wild edible mushroom *R. stricta* is thought to be an important antioxidant source. It is recommended that daily consumption is limited in terms of element toxicity.

REFERENCES

- Acharya, K., Das, K., Paloi, S., Dutta, A.K., Hembrom, M.E., Khatua, S., Parihar, A. (2017). Exploring a novel edible mushroom *Ramaria subalpina*: Chemical characterization and Antioxidant activity. *Pharmacognosy Journal*, 9, 30-34
- Aprotosoae, A.C., Zavastin, D.E., Mihai, C.T., Voichita, G., Gherghel, D., Sillion, M., Trifan, A., Miron, A. (2017). Antioxidant and antigenotoxic potential of *Ramaria lagentii* Marr & DE Stuntz, a wild edible mushroom collected from Northeast Romania. *Food and Chemical Toxicology*, 108, 429-437.
- Baba, H., Ergün, N., Özçubukçu, S. (2012). Antakya (Hatay)'dan toplanan bazı makrofungus türlerinde ağır metal birikimi ve mineral tayini. *Biyoloji Bilimleri Araştırma Dergisi*, 5(1), 5-6.
- Bae, M.J., Kim, K.J., Kim, S.J., Ye, E.J. (2007). Effect of mycelia extracts from *Lentinus edodes* mushroom-cultured *Astragalus membranaceus* Bunge on anti-cancer and anti-allergy activities. *Journal of the Korean Society of Food Science and Nutrition*, 36(1), 8-13.
- Bakır, K.T., Boufars, M., Karadeniz, M., Ünal, S. (2018). Amino acid composition and antioxidant properties of five edible mushroom species from Kastamonu, Turkey. *African Journal of Traditional, Complementary and Alternative Medicines*, 15, 80-87.
- Bal, C., Akgül, H., Sevindik, M., Akata, I., Yumrutas, O. (2017). Determination of the anti-oxidative activities of six mushrooms. *Fresen. Environ. Bull*, 26, 6246-6252.
- Bal, C., Sevindik, M., Akgül, H., Selamoğlu, Z. (2019). Oxidative Stress index and Antioxidant Capacity of *Lepista nuda* Collected from Gaziantep/Turkey. *Sigma*, 37(1), 1-5.
- Bozdoğan, A., Eker, T., Bozok, F., Ulukanlı, Z., Dogan, H.H., Buyukalaca, S. (2016). Multiple Antioxidant and Bioherbicidal Assays of the Edible Mushroom Species *Ramaria flava* in the Amanos Mountains. *Biointerface Research in Applied Chemistry*, 6, 1681-1685.
- Canlı, K., Altuner, E. M., Akata, I., Turkmen, Y., Uzek, U. (2016). In vitro antimicrobial screening of *Lycoperdon lividum* and determination of the ethanol extract composition by gas chromatography/mass spectrometry. *Bangladesh Journal of Pharmacology*, 11(2), 389-394.
- de Oliveira, J.M., Jordao, B.Q., Ribeiro, L.R., da Eira, A.F., Mantovani, M.S. (2002). Anti-genotoxic effect of aqueous extracts of sun mushroom (*Agaricus blazei* Murill lineage 99/26) in mammalian cells in vitro. *Food and Chemical Toxicology*, 40(12), 1775-1780.
- Devi, K.S.P., Maiti, K.T. (2016). Immunomodulatory and anti-cancer properties of pharmacologically relevant mushroom glycans. *Recent patents on biotechnology*, 10(1), 72-78.
- El Dine, R.S., El Halawany, A.M., Ma, C.M., Hattori, M. (2008). Anti-HIV-1 protease activity of lanostane triterpenes from the vietnamese mushroom *Ganoderma colossum*. *Journal of natural products*, 71(6), 1022-1026.
- Erel, O. (2004). A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clinical biochemistry*, 37(4): 277-285.
- Erel, O. (2005). A new automated colorimetric method for measuring total oxidant status. *Clinical biochemistry*, 38(12): 1103-1111.
- Falandysz, J., Kunito, T., Kubota, R., Bielawski, L., Mazur, A., Falandysz, J.J., Tanabe, S. (2007). Selected elements in brown birch scaber stalk *Leccinum scabrum*. *Journal of Environmental Science and Health A*, 42, 2081-2088.
- Falandysz, J., Kunito, T., Kubota, R., Gucia, M., Mazur, A., Falandysz, J.J., Tanabe, S. (2008). Some mineral constituents of parasol mushroom (*Macrolepiota procera*). *Journal of Environmental Science and Health B*, 43, 187-192.
- Froufe, H.J., Abreu, R.M.V., Ferreira, I.C. (2009). A QCAR model for predicting antioxidant activity of wild mushrooms. *SAR and QSAR in Environmental Research*, 20, 579-590.
- Gedik, G., Dülger, G., Asan, H., Özyurt, A., Allı, H., Asan, A. (2019). The antimicrobial effect of various formulations obtained from *Fomes fomentarius* against hospital isolates. *Mantar Dergisi*, 10(2), 103-109.
- Guo, Y.J., Deng, G.F., Xu, X.R., Wu, S., Li, S., Xia, E. Q., Li, F., Chen, F., Ling, W.H., Li, H.B. (2012). Antioxidant capacities, phenolic compounds and

- polysaccharide contents of 49 edible macrofungi. *Food & Function*, 3, 1195-1205.
- Gursoy, N., Sarikurkcu, C., Tepe, B., Solak, M.H. (2010). Evaluation of antioxidant activities of 3 edible mushrooms: *Ramaria flava* (Schaef.: Fr.) Quél., *Rhizopogon roseolus* (Corda) TM Fries., and *Russula delica* Fr. *Food Science and Biotechnology*, 19, 691-696.
- Gürgen, A., Yildiz, S., Can, Z., Tabbouche, S., Kiliç, A.O. (2018). Antioxidant, Antimicrobial and Anti-Quorum Sensing Activities of Some Wild and Cultivated Mushroom Species Collected from Trabzon, Turkey. *Fresen. Environ. Bull.* 27, 4120-4131.
- İnci, Ş., Kırbağ, S. (2018). *Terfezia clavaryi* Chatin'ın besinsel içeriği, antioksidan ve antimikrobiyal aktivitesi. *Artvin Çoruh Üniversitesi Orman Fakültesi Dergisi*, 19(2), 138-143.
- İnci, Ş., Dalkılıç, L.K., Dalkılıç, S., Kırbağ, S. (2019). *Helvella leucomelaena* (Pers.) Nannf.'ın antimikrobiyal ve antioksidan Etkisi. *Artvin Çoruh Üniversitesi Orman Fakültesi Dergisi*, 20(2), 249-253.
- Jose, N., Ajith, T.A., Janardhanan, K.K. (2002). Antioxidant, anti-inflammatory, and antitumor activities of culinary-medicinal mushroom *Pleurotus pufmonanus* (Fr.) Quel. (Agaricomycetidae). *International Journal of Medicinal Mushrooms*, 4(4).
- Kang, M.G., Bolormaa, Z., Lee, J.S., Seo, G.S., Lee, J.S. (2011). Antihypertensive activity and anti-gout activity of mushroom *Sarcodon aspratus*. *The Korean Journal of Mycology*, 39(1), 53-56.
- Khatua, S., Mitra, P., Chandra, S., Acharya, K. (2015). In vitro protective ability of *Ramaria aurea* against free radical and identification of main phenolic acids by HPLC. *Journal of herbs, spices & medicinal plants*, 21, 380-391.
- Kim, G.Y., Jeong, H.W., Jeong, D.J., Song, H.B., Lee, H.G. (2013). Effects of Shiitake Mushroom on Anti-platelet Aggregation and Anti-thrombotic. *Journal of Physiology & Pathology in Korean Medicine*, 27(2), 239-245.
- Kim, S.H., Song, Y.S., Kim, S.K., Kim, B.C., Lim, C.J., Park, E.H. (2004). Anti-inflammatory and related pharmacological activities of the n-BuOH subfraction of mushroom *Phellinus linteus*. *Journal of ethnopharmacology*, 93(1), 141-146.
- Korkmaz, A.I., Akgul, H., Sevindik, M., Selamoglu, Z. (2018). Study on determination of bioactive potentials of certain lichens. *Acta Alimentaria*, 47(1), 80-87.
- Kuo, M. (2009). *Ramaria stricta*. Retrieved from the Mushroom Expert. Com Web site: http://www.mushroomexpert.com/ramaria_stricta.html
- Lavi, I., Friese, D., Geresh, S., Hadar, Y., Schwartz, B. (2006). An aqueous polysaccharide extract from the edible mushroom *Pleurotus ostreatus* induces anti-proliferative and pro-apoptotic effects on HT-29 colon cancer cells. *Cancer letters*, 244(1), 61-70.
- Li, H.J., Chen, H.Y., Fan, L.L., Jiao, Z.H., Chen, Q.H., Jiao, Y.C. (2015). In vitro antioxidant activities and in vivo anti-hypoxic activity of the edible mushroom *Agaricus bisporus* (Lange) Sing. *Chaidam. Molecules*, 20(10), 17775-17788.
- Li, H. (2017). Extraction, purification, characterization and antioxidant activities of polysaccharides from *Ramaria botrytis* (Pers.) Ricken. *Chemistry Central Journal*, 11, 24.
- Liu, K., Wang, J., Zhao, L., Wang, Q. (2013). Anticancer, antioxidant and antibiotic activities of mushroom *Ramaria flava*. *Food and chemical toxicology*, 58, 375-380.
- Nguyen, T.K., Im, K.H., Choi, J., Shin, P.G., Lee, T.S. (2016). Evaluation of antioxidant, anti-cholinesterase, and anti-inflammatory effects of culinary mushroom *Pleurotus pulmonarius*. *Mycobiology*, 44(4), 291-301.
- Nikkarinen, M., Mertanen, E. (2004). Impact of geological origin on trace element composition of edible mushrooms. *Journal of Food Composition and Analysis*, 17(3-4), 301-310.
- Öztürk, M., Tel, G., Öztürk, F.A., Duru, M.E. (2014). The cooking effect on two edible mushrooms in Anatolia: fatty acid composition, total bioactive compounds, antioxidant and anticholinesterase activities. *Records of Natural Products*, 8, 189.
- Pandimeena, M., Prabu, M., Sumathy, R., Kumuthakalavalli, R. (2015). Evaluation of phytochemicals and *in vitro* anti-inflammatory, anti-diabetic activity of the white oyster mushroom, *Pleurotus florida*. *Int. Res. J. Pharmaceut. Appl. Sci*, 5, 16-21.
- Ramesh, C.H., Pattar, M.G. (2010). Antimicrobial properties, antioxidant activity and bioactive compounds from six wild edible mushrooms of western ghats of Karnataka, India. *Pharmacognosy research*, 2, 107.
- Rathore, H., Prasad, S., Sharma, S. (2017). Mushroom nutraceuticals for improved nutrition and better human health: A review. *Pharma Nutrition*, 5(2), 35-46.
- Sadi, G., Kaya, A., Yalcin, H.A., Emsen, B., Kocabas, A., Kartal, D.I., Altay, A. (2016). Wild edible mushrooms from Turkey as possible anticancer agents on HepG2 cells together with their antioxidant and antimicrobial properties. *International journal of medicinal mushrooms*, 18, 83-95
- Sarikurkcu, C., Copur, M., Yildiz, D., Akata, I. (2011). Metal concentration of wild edible mushrooms in Soguksu National Park in Turkey. *Food Chemistry*, 128(3), 731-734.
- Selamoglu, Z., Akgul, H., Dogan, H. (2016). Environmental effects on biologic activities of pollen samples obtained from different phytogeographical regions in Turkey. *Fresenius Environmental Bulletin*, 25, 2484-2489.
- Selamoglu, Z., Dugun, C., Akgul, H., Gulhan, M.F. (2017). In-vitro antioxidant activities of the ethanolic extracts of some contained-allantoin plants. *Iranian journal of pharmaceutical research: IJPR*, 16 (Suppl), 92.
- Sevindik, M. (2018). Investigation of antioxidant/oxidant status and antimicrobial activities of *Lentinus tigrinus*. *Advances in Pharmacological Sciences*, 2018. <https://doi.org/10.1155/2018/1718025>.

- Sevindik, M. (2019). The novel biological tests on various extracts of *Cerioporus varius*. *Fresenius Environmental Bulletin*, 28(5), 3713-3717.
- Sevindik, M., Akgul, H., Akata, I., Alli, H., Selamoglu, Z. (2017). *Fomitopsis pinicola* in healthful dietary approach and their therapeutic potentials. *Acta Alimentaria*, 46(4), 464-469.
- Sevindik, M., Akgul, H., Bal, C., Selamoglu, Z. (2018a). Phenolic contents, oxidant/antioxidant potential and heavy metal levels in *Cyclocybe cylindracea*. *Indian Journal of Pharmaceutical Education and Research*, 52(3), 437-441.
- Sevindik, M., Akgul, H., Dogan, M., Akata, I., Selamoglu, Z. (2018b). Determination of antioxidant, antimicrobial, DNA protective activity and heavy metals content of *Laetiporus sulphureus*. *Fresenius Environmental Bulletin*, 27(3), 1946-1952.
- Sevindik, M., Rasul, A., Hussain, G., Anwar, H., Zahoor, M.K., Sarfraz, I., Kamran, K.S., Akgul, H., Akata, I., Selamoglu, Z. (2018c). Determination of anti-oxidative, anti-microbial activity and heavy metal contents of *Leucoagaricus leucothites*. *Pak. J. Pharm. Sci*, 31(5), 2163-2168.
- Sharma, S.K., Gautam, N. (2017). Chemical and Bioactive Profiling, and Biological Activities of Coral Fungi from Northwestern Himalayas. *Scientific reports*, 7, 46570.
- Song, Y.S., Kim, S.H., Sa, J.H., Jin, C., Lim, C.J., Park, E.H. (2003). Anti-angiogenic, antioxidant and xanthine oxidase inhibition activities of the mushroom *Phellinus linteus*. *Journal of Ethnopharmacology*, 88(1), 113-116.
- Vetter, J. (1990). Mineral element content of edible and poisonous macrofungi. *Acta Alimentaria*, 19(1), 27-40.
- Weng, Y., Xiang, L., Matsuura, A., Zhang, Y., Huang, Q., Qi, J. (2010). Ganodermasides A and B, two novel anti-aging ergosterols from spores of a medicinal mushroom *Ganoderma lucidum* on yeast via UTH1 gene. *Bioorganic & Medicinal Chemistry*, 18(3), 999-1002.
- Zengin, G., Uren, M.C., Kocak, M.S., Gungor, H., Locatelli, M., Aktumsek, A., Sarikurkcu, C. (2017). Antioxidant and enzyme inhibitory activities of extracts from wild mushroom species from Turkey. *International Journal of Medicinal Mushrooms*, 19, 327-336.
- ***Institute of Medicine. Food and Nutrition Board. (2001). Dietary reference intakes for vitamin A, vitamin K, arsenic, boron, chromium, copper, iodine, iron, manganese, molybdenum, nickel, silicon, vanadium, and zinc. A Report of the Panel on Micronutrients, Subcommittees on Upper Reference Levels of Nutrients and of Interpretation and Uses of Dietary Reference Intakes, and the Standing Committee on the Scientific Evaluation of Dietary Reference Intakes Food and Nutrition Board Institute of Medicine.
- ***WHO (2000). Nickel. Chapter 6.10. Air Quality Guidelines - Second Edition. WHO Regional Office for Europe, Copenhagen, Denmark. 1-17.
- ***WHO (2001). Lead. Chapter 6.7. Air Quality Guidelines - Second Edition. WHO Regional Office for Europe, Copenhagen, Denmark. 1-17.